

# Guadial A and Psiguadials C and D, Three Unusual Meroterpenoids from *Psidium guajava*

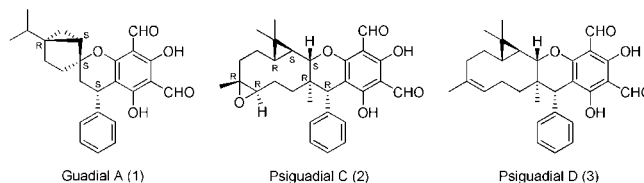
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## ABSTRACT

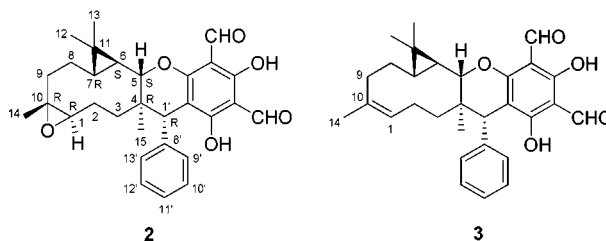
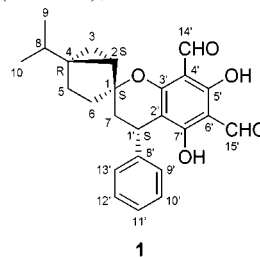


The first monoterpene-based meroterpenoid (1) and two novel sesquiterpene-based ones (2 and 3) with unprecedented skeletons were isolated from the leaves of *Psidium guajava*. Their structures with absolute configuration were elucidated by extensive spectroscopic studies. A plausible biosynthetic pathway for all meroterpenoids from the title plant is also proposed. Compounds 2 and 3 showed significant cytotoxicity toward HepG2 and HepG2/ADM cells.

The leaves of *Psidium guajava* L. (Myrtaceae) have long been used in China as folk medicine for the treatment of diarrhea and hyperglycemia. In recent years, several sesquiterpene-based meroterpenoids with unprecedented skeletons have been isolated from the leaves of *P. guajava* by several research groups.<sup>1,2</sup> Some of the *Psidium* meroterpenoids exhibited significant biological activities including inhibitory effects on protein tyrosine phosphatase 1B (PTP1B) and the growth of human hepatoma cell (HepG2).<sup>1,2</sup> The main structural difference of these sesquiterpene-based *Psidium* meroterpenoids could be attributed to the skeleton of the terpenoid moiety as well as the coupling pattern between benzyl phloroglucinol dialdehyde and terpenoid. Two plausible

biosynthetic pathways and a short biomimetic synthesis relative to *Psidium* meroterpenoids had been also reported.<sup>1,3</sup> However, there is still confusion about their biogenetic pathway.

As a result of our continuing study on this plant, the first monoterpene-based meroterpenoid, guadial A (1), and two novel sesquiterpene-based meroterpenoids, psiguadials C and D (2 and 3), were isolated. In this paper, the



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biogenetic route of all meroterpenoids from the title plant and cytotoxic effects of **1**–**3** were also described.

Guadial A (**1**) was isolated as a colorless oil. The molecular formula was established as  $C_{25}H_{26}O_5$  by HR-ESI-MS at  $m/z$  429.1671 [ $M + Na$ ]<sup>+</sup> (calcd for  $C_{25}H_{26}O_5Na$  429.1673). The UV spectrum of **1** showed the absorptions maxima at 284 and 302 nm. The IR spectrum exhibited the characteristic absorptions for hydroxyl ( $3500\text{ cm}^{-1}$ ), aldehyde ( $2927\text{ cm}^{-1}$ ), and benzene ring ( $1632, 1436, \text{ and } 697\text{ cm}^{-1}$ ). The  $^1H$  and  $^{13}C$  NMR spectra displayed the presence of two chelated phenolic hydroxyls, two aldehydes, a monosubstituted benzene ring, a hexasubstituted benzene ring, and two methyls. The above spectral data suggested that **1** was a benzylphloroglucinol dialdehyde–terpene derivative.<sup>1,2</sup> Detailed analysis of  $^1H$ – $^1H$  COSY, HSQC, and HMBC spectra resulted in unambiguous assignment of all  $^1H$  and  $^{13}C$  NMR signals as shown in Table 1.

The  $^1H$ – $^1H$  COSY spectrum showed the presence of five spin coupling systems in bold as shown in Figure 1. In the HMBC spectrum, the correlations between H<sub>3</sub>-9/H<sub>3</sub>-10 and C-4, between H-2/H<sub>2</sub>-7 and C-6, between H<sub>2</sub>-3 and C-5, and between H<sub>2</sub>-3/H<sub>2</sub>-5 and C-8 proved the presence of a monoterpenoid moiety with a 3/5 bicyclic ring system and an isopropyl group at C-4 (**1a**). Furthermore, the HMBC correlations between H-5 and C-1, and between H<sub>2</sub>-7 and C-2/C-6 established an oxa-spiro[5.5] ring. Based on the molecular formula information, the remaining oxygen atom was assigned to bridge C-1 and C-3'. Comparison of the NMR data of **1** with those of euglobal-Ib<sup>4</sup> suggested that their structures were very similar except that the isopropyl group at C-1' in euglobal-Ib was replaced by a monosubstituted benzene ring in **1**.

The relative configuration of **1** could be deduced by its ROESY spectrum. In the ROESY spectrum, the correlations between H-3a and H-6a, between H-3b and H-8, between H-2 and H-1', between H-7a and H-6b, as well as between H-7b and H-2 indicated that these protons were cofacial, respectively. Thus, the relative configuration of **1** was established as shown in Figure 2.

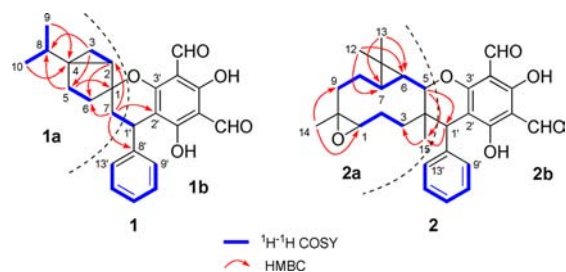
The absolute configuration of **1** was elucidated by quantum chemical CD calculation.<sup>2,5</sup> The overall predicted CD spectra of (1*S*,2*S*,4*R*,1'*S*)-**1** and (1*R*,2*R*,4*S*,1'*R*)-**1** were compared with the experimental one (Figure 3). The result showed that the spectrum calculated for (1*S*,2*S*,4*R*,1'*S*)-**1** was consistent with the experimental one. Therefore, the absolute configuration of **1** was determined as 1*S*,2*S*,4*R*,1'*S*.

Psiguadial C (**2**) was obtained as colorless needles. The molecular formula was assigned as  $C_{30}H_{34}O_6$  by its HR-EI-MS ( $m/z$  490.2352 [ $M$ ]<sup>+</sup>, calcd for  $C_{30}H_{34}O_6$  490.2350). The UV and IR spectra of **2** showed the characteristic absorptions for *Psidium* meroterpenoids.<sup>1,2</sup> The NMR spectra of **2** also displayed the signals for benzylphloroglucinol dialdehyde and sesquiterpenoid moieties. With the aid of  $^1H$ – $^1H$  COSY, HSQC, and

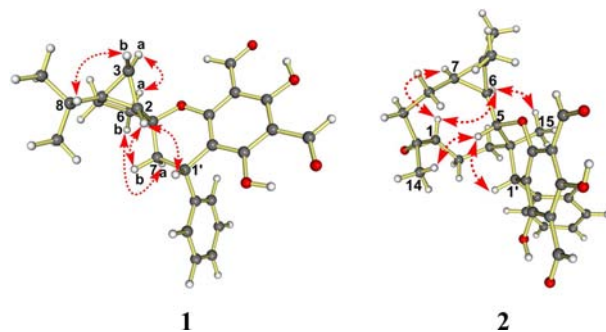
**Table 1.** NMR Data of **1** (in  $CDCl_3$ ,  $J$  in Hz)<sup>a</sup>

no.	$\delta_H$	$\delta_C$	no.	$\delta_H$	$\delta_C$
1		88.9	1'	4.21 (dd, 7.3, 9.5)	35.1
2	1.35 (dd, 3.6, 7.8)	28.3	2'		103.7
3	a 0.82 (dd, 3.6, 5.2) b 0.45 (dd, 5.2, 7.8)	12.2	3'		165.9
			4'		104.6
4		34.6	5'		168.6
5	1.67 (m)	24.5	6'		104.3
6	a 1.70 (m) b 1.64 (m)	33.6	7'		169.9
			8'		144.7
7	a 2.01 (dd, 9.5, 14.3) b 2.34 (dd, 7.3, 14.3)	42.3	9'	7.15 (br d, 7.0)	126.8
			10'	7.29 (br d, 7.0)	128.7
8	1.37 (m)	32.6	11'	7.21 (t, 7.0)	126.5
9	0.97 (d, 6.8)	19.8	12'	7.29 (br d, 7.0)	128.7
10	0.91 (d, 6.8)	19.7	13'	7.15 (br d, 7.0)	126.8
5'-OH	13.53 (s)		14'	10.10 (s)	192.5
7'-OH	13.17 (s)		15'	10.13 (s)	191.8

<sup>a</sup> Overlapped signals were reported without designating multiplicity.



**Figure 1.** Key  $^1H$ – $^1H$  COSY and HMBC correlations of **1** and **2**.



**Figure 2.** Key ROESY correlations of **1** and **2**.

HMBC experiments, all of the  $^1H$  and  $^{13}C$  NMR signals of **2** were assigned as shown in Table 2.

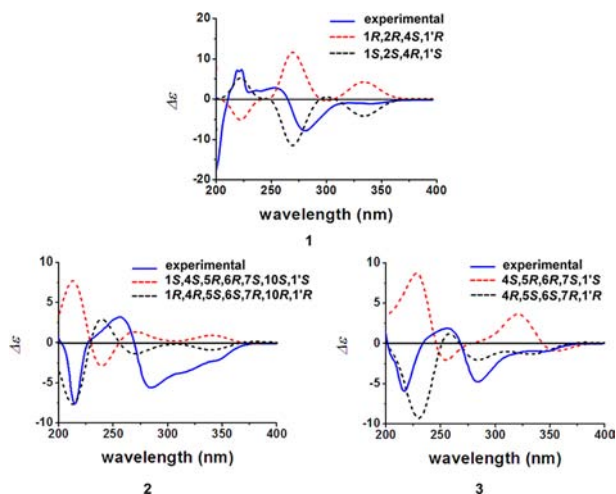
The presence of two spin systems of H-1–H-3 and H-5–H-9 in the  $^1H$ – $^1H$  COSY spectrum, as well as the HMBC correlations between H-5 and C-15, between H-15 and C-3, and between H-14 and C-1/C-9 suggested the existence of a 10-membered carbon ring in **2** (Figure 1). The HMBC correlations between H-12/H-13 and C-6/C-7 indicated the presence of a cyclopropane ring between

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**Figure 3.** Calculated and experimental CD spectra of **1**–**3**.

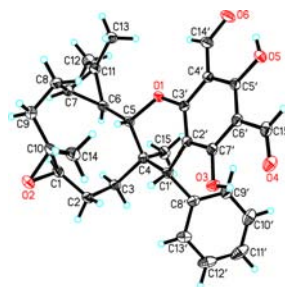
C-6 and C-7. The above data indicated the presence of a sesquiterpenoid moiety with 3/10 consecutive carbocyclic system (**2a**). The HMBC correlations between H-1' and C-15 and between H-5 and C-1' (Figure 1) indicated that the sesquiterpenoid moiety (**2a**) and benzylphloroglucinol dialdehyde (**2b**) were connected via a C-4–C-1' bond. Moreover, the obvious upfield chemical shifts of C-3' ( $\delta_C$  165.8) and the oxygenated methine C-5 ( $\delta_C$  84.0) suggested the oxygen atom was bridged C-5 and C-3' to form a dihydropyran ring.

The relative stereochemistry of **2** could be elucidated by a ROESY experiment. The NOE correlations between H-5 and H-14/H-1', between H-1 and H-6/H-7, and between H-15 and H-6 established the relative configuration of **2** as shown in Figure 2.

The structure and stereochemistry of **2** were further confirmed by an X-ray experiment. In the X-ray structure of **2** (Figure 4), the 10-membered ring was *trans*-fused with the dihydropyran and epoxide rings but *cis*-fused with the cyclopropane ring. The final refinement on the CuK $\alpha$  data resulted in a Flack parameter of 0.01(13), allowing an unambiguous assignment of the absolute structure of **2** (1*R*,4*R*,5*S*,6*S*,7*R*,10*R*,1'*R*), which was consistent with the result obtained by the following quantum chemical CD calculation experiment (Figure 3).

The molecular formula of psiguadial D (**3**) was established to be C<sub>30</sub>H<sub>34</sub>O<sub>5</sub> by its HR-EI-MS at  $m/z$  474.2395 [M]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>34</sub>O<sub>5</sub> 474.2406). Comparison of the <sup>1</sup>H and <sup>13</sup>C NMR data of **3** with those of **2** revealed that they were very similar except for the presence of an olefinic group ( $\delta_H$  5.31;  $\delta_C$  127.5 and 130.7) in **3** instead of the epoxide unit in **2**. In the HMBC spectrum, correlations between H-1 and C-9/C-14 indicated that the double bond was located at C-1 and C-10. Finally, the absolute configuration of **3** was established to be 4*R*,5*S*,6*S*,7*R*,1'*R* by quantum chemical CD calculation experiment (Figure 3).

To date, nine meroterpenoids had been isolated from *P. guajava*.<sup>1,2</sup> Two plausible biosynthetic pathways relative to these *Psidium* meroterpenoids have also been reported.<sup>1</sup>



**Figure 4.** X-ray structure of **2**.

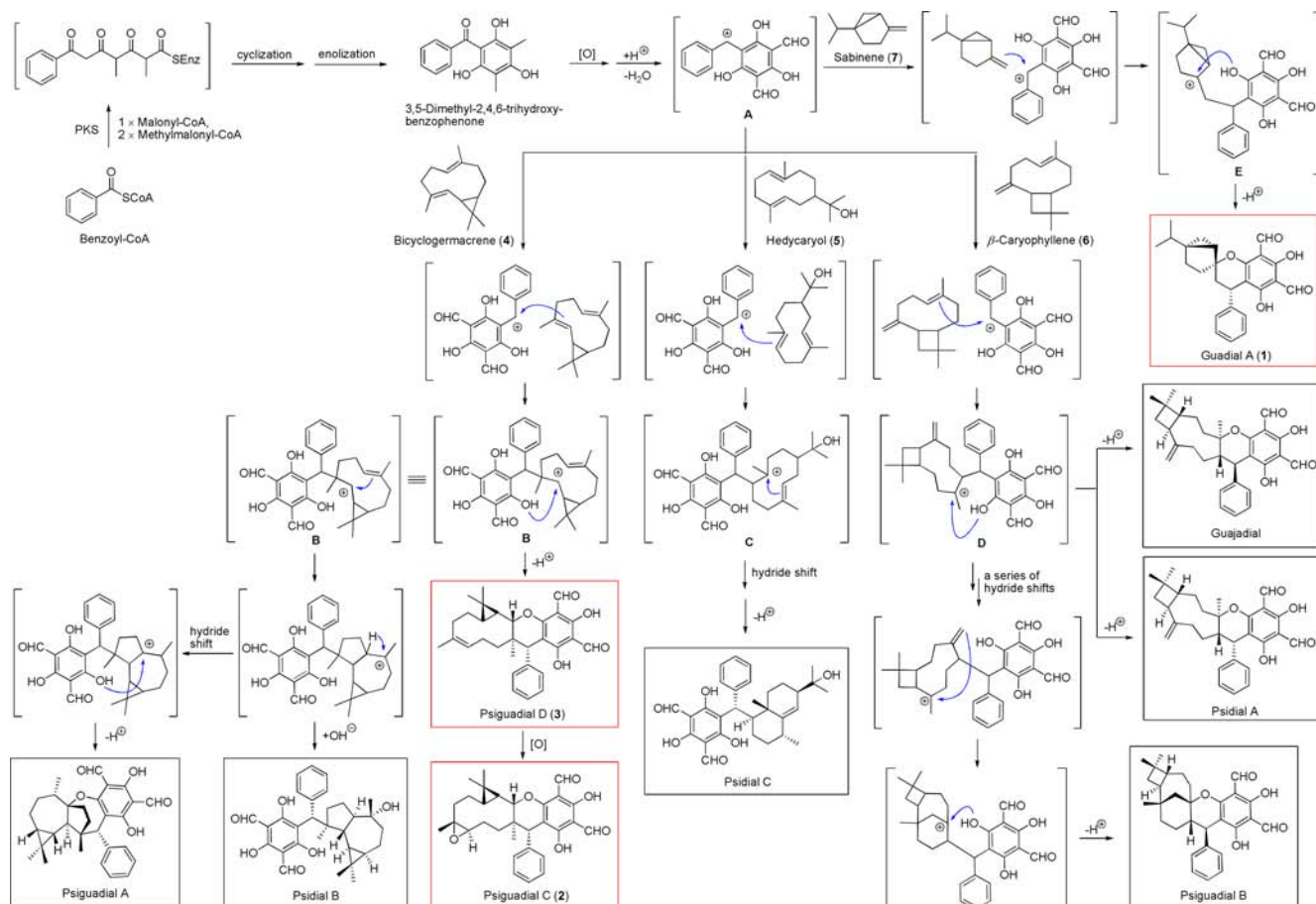
**Table 2.** NMR Data of **2** and **3** (in CDCl<sub>3</sub>,  $J$  in Hz)<sup>a</sup>

no.	<b>2</b>		<b>3</b>	
	$\delta_H$	$\delta_C$	$\delta_H$	$\delta_C$
1	2.93 (dd, 7.2, 1.2)	64.0	5.31 (dd, 10.4, 3.3)	127.5
2	a 2.11 (dd, 7.2, 14.9) b 1.84 (m)	22.5	a 2.76 (m) b 2.10 (m)	23.4
3	1.68	31.6	1.57	35.4
4		40.6		41.4
5	3.85 (d, 6.7)	84.0	3.71 (d, 7.1)	85.3
6	0.98 (m)	26.6	0.95	27.0
7	0.67 (m)	31.7	0.66 (td, 1.4, 10.4)	31.6
8	a 2.03 (dd, 5.5, 13.9) b 1.16	21.3	a 1.92 b 0.96	22.4
9	a 2.21 (dd, 5.4, 12.4) b 1.17	39.3	2.09	38.1
10		60.4		130.7
11		21.6		19.8
12	1.20 (s)	19.2	1.20 (s)	30.3
13	1.20 (s)	30.3	1.13 (s)	19.3
14	1.34 (s)	17.2	1.72 (s)	17.5
15	0.77 (s)	18.5	0.73 (s)	18.6
1'	4.23 (s)	43.9	4.39 (s)	43.9
2'		104.6		105.2
3'		165.8		166.4
4'		104.3		104.5
5'		168.3		168.3
6'		104.8		104.2
7'		170.6		170.6
8'		139.7		140.4
9'	6.79 (d, 7.7)	127.6	6.78 (d, 7.8)	127.5
10'	7.15	127.4	7.15	127.5
11'	7.23	126.5	7.23	126.3
12'	7.19	130.1	7.33	130.2
13'	7.31	127.7	7.33	127.8
14'	10.13 (s)	192.1	10.12 (s)	192.2
15'	10.13 (s)	191.7	10.10 (s)	191.6
5'-OH	13.60 (s)		13.60 (s)	
7'-OH	13.15 (s)		13.14 (s)	

<sup>a</sup> Overlapped signals are reported without designating multiplicity.

Because of the findings of key biogenetic intermediate and terpene precursors in this plant, the plausible biogenetic route of *Psidium* meroterpenoids could be proposed as shown in Scheme 1. First, the benzoyl-CoA was successively condensed with one molecule of malonyl-CoA and

**Scheme 1.** Plausible Biosynthetic Pathway for *Psidium* Meroterpenoids from *P. guajava*



two molecules of methylmalonyl-CoA. Then, cyclization and enolization should lead to the formation of intermediate 3,5-dimethyl-2,4,6-trihydroxybenzophenone, which had been previously isolated from the same plant.<sup>6</sup> The intermediate could be oxidized and then generated a carbocation **A**. As a cationic initiator, the carbocation **A** could respectively couple with different terpene precursors including bicyclgermacrene (**4**), hedycaryol (**5**),  $\beta$ -caryophyllene (**6**), and sabinene (**7**), which had been detected in the plant (see the Supporting Information), to generate carbocations **B–E**. Further rearrangement of carbocations **B–E** could construct different meroterpenoid skeletons to afford guajadial,<sup>1a</sup> psidials A–C,<sup>1b</sup> psiguadials A and B,<sup>2</sup> as well as compounds **1–3**.

The cytotoxic effects of **1–3** were evaluated by MTT assay in doxorubicin-sensitive and -resistant human hepatoma cells (HepG2 and HepG2/ADM) as described previously.<sup>2</sup> Compounds **2** and **3** exhibited potent inhibitory effects on the growth of HepG2 cells with  $IC_{50}$  values of  $104.5 \pm 13.71$  nM and  $128.3 \pm 18.2$  nM, respectively. The cytotoxicity of **2** and **3** in HepG2/ADM, with  $IC_{50}$  values of  $21.06 \pm 1.25$   $\mu$ M and  $23.65 \pm 1.71$   $\mu$ M, differed significantly from that in HepG2 cells. As with other

*Psidium* meroterpenoids,<sup>2</sup> **2** and **3** might also be substrates for the efflux transporter P-glycoprotein pump.

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**Supporting Information Available.** Detailed description of the experimental procedure, a listing of UV, IR, HR-EI-MS, and NMR spectra of compounds **1–3**, CIF files of **2**, quantum chemical CD calculations of **1–3**, and GC-MS analysis of **4–7**. These materials are available free of charge via the Internet at <http://pubs.acs.org>.

The authors declare no competing financial interest.